

Research Article

Proteomic Analysis and Characterization of Amylin (IAPP) in *Homo Sapiens*

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Abstract

Amylin which is also called as Islet amyloids polypeptide (IAPP), is a pancreatic B-cell hormone co-released with insulin. Amyloids are insoluble fiber and the abnormal accumulation of these fibers in the tissue of the pancreas leads to type II diabetes. Bioinformatics tools were applied to process, prioritise and annotate the raw sequence which is then converted into meaningful information about the proteins. These methods are useful in the medical research as they increase the annotation of proteome through functional and structural genomic efforts. Firstly, the result of primary structure analysis reveals that most of the amylin are hydrophobic in nature due to presence of high non-polar residues content. Amylin has both acidic and basic property as its isoelectric point ranged from 5.31-9.83. The aliphatic index computed by ExPasy's ProtParam infers that most of the amylin are stable at wide range of temperature (66.84-108.43). The secondary structure analysis by SOPMA reveals that amylin contain more random coils and beta sheets. SMART values revealed an expected range of 0.0000198-27. The Ligand binding site of an amylin was obtained by SWISS-MODEL as it determines the 3D structure of a protein. The RNA structure was predicted by using Genebee service software.

Keywords: IAPP, Type 2 Diabetes, ExPasy ProtParam, SOPMA, SMART, SWISS-MODEL.

Introduction

Amylin is a protein which is insoluble in low tissue concentration. The formation of amyloids, is known to cause non-neuronal diseases such as Diabetes mellitus type II. Amylin forms long stable fibres which are arranged in regular pattern [Nanga *et al*, 2008]. Islet amyloid polypeptide (IAPP, also called amylin) is associated with type II diabetes, a disease that has maximized over the past two decades and now afflicts an estimated 20 million people in America and 100 million worldwide [Hossain *et al*, 2007]. The role of IAPP in diabetes is uncertain [Hoppener *et al*, 2002], but there is escalating evidence for the consequence of amyloid formation associated with this disease. Ninety-five percent of analysed individuals stain positive posthumous for pancreatic amyloid deposits composed of mature, fibrillar IAPP [Hoppener *et al*, 2000]. Gene responsible for the formation of amyloid is located on the 12p12.1 chromosome. Pancreatic extracts from respective person with type II diabetes have been found to contain extracellular deposits of the 37-residue peptide hormone IAPP that is expressed, stored, and secreted from the beta-islet cells of the pancreas [Cooper, 1994; Cooper *et al*, 1987]. IAPP was first identified in 1987, and was considered as a

Potential target for the treatment of diabetes [Schmitz *et al*, 2004; Yan *et al*, 2006]. Amyloids are insoluble fibrillar aggregates of proteins / peptides. Type-2 diabetes mellitus (DM-2) was one such disease associated with human islet amyloid polypeptide (hIAPP, also called amylin), which is a peptide of 37 residues secreted from beta-cells of the pancreas. hIAPP (Human Islet Amyloid Polypeptide) is monomeric in its physiological state but aggregated in the disease state. Evidence from the cell and animal studies indicates a link between hIAPP misfolding and pancreatic beta cell dysfunction. [Balali-Mood *et al*, 2005; Cooper *et al*, 1987; Eisenhaber *et al*, 1996; Gill and Von Hippel, 1989]. The misfolding or abnormal accumulation of these peptides in the tissue of the pancreas leads to the metabolic disorder DM-2 [Zanuy *et al*, 2003]. Misfolding or unfolding of native protein exposes hydrophobic regions which results in unstable intermediates of revised conformation that have the propensity to form oligomers, where these oligomers form pathogenic subunits and crossed beta sheets. The soluble oligomers become toxic and accumulate to promote apoptosis [Balali-Mood *et al*, 2005].

Researchers are trying to find out the aberrant metabolic conditions and factors which can possibly trigger or can have the potential to hinder amyloid formation. Some evidence suggests that altered hIAPP

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sequences and abnormal metabolic conditions might be responsible for development of this disease [Hayden, 2002]. Human amylin exhibits physicochemical properties predisposing the peptide hormone to accumulate and form Amyloids fibers, which may play a part in B-cell destruction in type 2 diabetes.

Several peptide segments derived from the amino acid sequence of IAPP are capable of composing amyloid-like fibrils, with earlier work suggesting that the segment containing residues 22–29 forms the spine of the fibril under physiological conditions [Westermark *et al*, 1990; Moriarty and Raleigh, 1999; Goldsbury *et al*, 2000; Tenidis *et al*, 2000.]. Amylin is a pancreatic B-cell hormone co-released with insulin in response to food intake [Young and Denaro, 1998].

The wealth of amylin protein sequence information that has been made publicly available in recent years requires the advancement of high-throughput functional genomics and proteomics approaches for its analysis. Such approaches need suitable data assimilation procedures and a high level of annotation in order to gain maximum benefit from the results generated. Identification of proteins of interest from a particular biological study requires the application of bioinformatics tools to process and compute the data. From a protein function standpoint, transfer of annotation from known proteins to a novel destination is currently the only practical way to convert vast quantities of raw sequence data into meaningful information. Advanced bioinformatics tools now provide more sophisticated methods to transfer functional annotation, consolidating sequence, family profile and structural search methodology. The importance of these approaches to medical research is growing as we move to annotate the proteome through functional and structural genomic efforts.

Detailed knowledge of amylin and their properties can be extensively studied through the physicochemical and the structural properties of the proteins are well figured out with the use of computational tools. The statistics about the sequence of amylin protein such as number of amino acid, frequency is predicted by CLC work bench (<http://www.clc.bio.com/index.php?id=28>). Modular Architecture Research Tool (SMART) is a biological database that is used in the identification and analysis of protein domains within protein sequences [Letunic *et al*, 2009; Schultz *et al*, 1998]. Sequence length, and the physico-chemical properties of a amylin protein such as molecular weight, atomic composition, extinction coefficient, GRAVY, aliphatic index, instability index, etc. can be computed by ProtParam. The TMpred program makes a prediction of membrane-spanning regions and their orientation. The algorithm was based on the statistical analysis of TMbase, a database of naturally occurring trans-membrane proteins. The prediction was made using a combination of several weight-matrices for scoring [Hofmann and Stoffel, 1993]. MUSCLE stands

for MULTiple Sequence Comparison by Log-Expectation. MUSCLE is claimed to achieve both better average accuracy and a better speed than ClustalW2 or T-Coffee. The protein 3D model and its characteristics can be predicted by Swiss model server [Tsetlin and Hucho, 2004]. Reverse Translate accepts a protein sequence as input and uses a codon usage table to achieve a DNA sequence representing the most likely non-degenerate coding sequence. A consent sequence derived from all the possible codons for each amino acid is also returned. Further Computer-aided techniques for the productive identification and optimization of novel molecules with a desired biological activity have become a part of the drug discovery process.

The raw sequence information of all the proteins and nucleic acid can convert to analytical and relative information with the help of soft computing tools [Ashokan and Pillai, 2008]. Prediction of protein function is important application of bioinformatics [Prashanth *et al*, 2010]. The amino acid sequence provides most of the information required for determining and characterizing the molecule's function, physical and chemical properties. Sequence analysis and physicochemical characterization of proteins using biocomputation tool have been done by many researchers and reported. [Ashokan *et al*, 2011; Madhu and Mahesh, 2015; Mahesh *et al*, 2015; Praveen and Mahesh, 2015; Vishwanath *et al*, 2015; Mandeep Singh *et al*, 2016; Amrutha *et al*, 2015].

The main objective of this study is to determine the physicochemical characterization of the amyloid protein and to predict its function. As Amyloids are associated with type 2 diabetes, characterization of Amylin can be used in the treatment of type 2 diabetes.

Materials and methods

Protein sequence retrieval: The Protein Sequences of amylin (10 sequences) were retrieved in FASTA format from NCBI database (Table 1).

Amino acid Composition: The amino acid composition of selected proteins were computed using the tool CLC free workbench (www.clc.bio.com/.../clc-main-workbench), tabulated in (Table 2).

Primary structure analysis: Counts of hydrophobic and hydrophilic residues were calculated from the primary structure analysis by CLC workbench (Table 3).

Physico-chemical parameters: The physicochemical parameters such as theoretical isoelectric point (Ip), molecular weight, total number of positive and negative residues, extinction coefficient, instability index [Gill and Von Hippel, 1989] aliphatic index [Eisenhaber *et al*, 1996] and grand average hydropathy (GRAVY) [Kitchen *et al*, 2007] were computed using the Expasy's ProtParam server [Mugilan *et al*, 2010], and tabulated in (Table 4).

Secondary structure prediction: The secondary structure was predicted by self-optimized prediction method with alignment by SOPMA server (<http://jing.cz3.nus.edu.sg/cgi-bin/svmprot.cgi>) [Ashokan *et al*, 2011] (Table 5).

Table1: Selected AMYLIN protein sequences retrieved from NCBI

| SI No | Species | ID | Length | Protein Sequence |
|-------|---------------------|-------------|--------|--|
| 1 | <i>Homo sapiens</i> | AAA35524.1 | 62aa | HQVEKRKNTATCATQRLANFLVHSSNFGAILSSTNVGSNTYGKRNAVEV LKREPLNYLPL |
| 2 | <i>Homo sapiens</i> | NP_000406.1 | 89aa | MGILKLQVFLIVLSVALNHLKATPIESHQVEKRKNTATCATQRLANFLVHS SNNFGAILSSTNVGSNTYGKRNAVEVLKREPLNYLPL |
| 3 | <i>Homo sapiens</i> | 3HGZ E | 37aa | KCNTATCATQRLANFLVHSSNFGAILSSTNVGSNTY |
| 4 | <i>Homo sapiens</i> | 2KB8 A | 37aa | KCNTATCATQRLANFLVHSSNFGAILSSTNVGSNTX |
| 5 | <i>Homo sapiens</i> | 2L86 A | 38aa | KCNTATCATQRLANFLVHSSNFGAILSSTNVGSNTYX |
| 6 | <i>Homo sapiens</i> | AAC50301.1 | 427aa | MMDAQYKCYDRMQQLPAYQGEPCYCNRTWDGWLCWDDTPAGVLSYQFC PDYFPDFDPSEKVTKYCGVWFKHPENNRTWSNYTMCNAFTPEKLNAYV LYYLAIVGHLSIFTLVISLGIFVFRSLGCRVTLHKNNFLTLYLNSMIIHHLV EVVPPNGELVRRDPVSKILHFFHQYMMACNYFWMLCEGIYHLTLIVAVFT EKQRLRWYLLGWGFLVPTTTHAITRAVYFNDNCWLSVETHLLYIHGVP MAALVVNFFLLNIVRVLTKMRETHEAESMYLKAVKATMTLVPLLGIF VVPWPRPSNKMLGKIYDVMHSLIHFQGFVATIYFCNNEVQTTVKRQW AQFKIQWNRWGRRPSNRSARAAAAAAEAGDIPYICHQELRNEPANNQG EESAEIIPLNIEQESSA |
| 7 | <i>Homo sapiens</i> | AAC50300.1 | 474aa | YCNRTWDGWLCWDDTPAGVLSYQFCPDYFPDFDPSEKVTKYCDEKGVWF KHPENNRTWSNYTMCNAFTPEKLNAYVLYYLAIVGHLSIFTLVISLGIFV FRSLGCRVTLHKNNFLTLYLNSMIIHHLVEVVPNGELVRRDPVSKILHFF HQYMMACNYFWMLCEGIYHLTLIVAVFTKQRLRWYLLGWGFLVPTT IHAITRAVYFNDNCWLSVETHLLYIHGVPMAALVVNFFLLNIVRVLTKM RETHEAESMYLKAVKATMILVPLLGIFVVPWPRPSNKMLGKIYDVMH SLIHFQGFVATIYFCNNEVQTTVKRQWAQFKIQWNRWGRRPSNRSARA AAAAAAEAGDIPYICHQEPNEPANNQGEESAEIIPLNIEQESSA |
| 8 | <i>Homo sapiens</i> | NP_001733.1 | 474aa | MRFTFTRCLALFLLNHPPTILPAFSNQTYPTIEPKFLYVVRGKKMMDAQ YKCYDRMQQLPAYQGEPCYCNRTWDGWLCWDDTPAGVLSYQFCPDYFPD FDPSEKVTKYCDEKGVWFKHPENNRTWSNYTMCNAFTPEKLNAYVLYYL AIVGHLSIFTLVISLGIFVFRSLGCRVTLHKNNFLTLYLNSMIIHHLVEVVP NGELVRRDPVSKILHFFHQYMMACNYFWMLCEGIYHLTLIVAVFTKQRL LRWYLLGWGFLVPTTTHAITRAVYFNDNCWLSVETHLLYIHGVPMAAL VVNFFLLNIVRVLTKMRETHEAESMYLKAVKATMILVPLLGIFVVP WRPSNKMLGKIYDVMHSLIHFQGFVATIYFCNNEVQTTVKRQWAQFKI QWNRWGRRPSNRSARAAAAAAEAGDIPYICHQELRNEPANNQGEESAEI IPLNIEQESSA |
| 9 | <i>Homo sapiens</i> | 2WK3 D | 42aa | DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVVIA |
| 10 | <i>Homo sapiens</i> | 2LMP K | 40aa | DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVV |

Table 2: Representation of frequency of amino acids in amylin

| Amino acid | AAA35524.1 | NP_000406.1 | 3HGZ E | 2KB8 A | 2L86 A | AAC50301.1 | AAC50300.1 | NP_001733.1 | 2WK3 D | 2LMP K |
|-------------------|------------|-------------|--------|--------|--------|------------|------------|-------------|--------|--------|
| Alanine (A) | 0.081 | 0.079 | 0.108 | 0.108 | 0.105 | 0.066 | 0.063 | 0.063 | 0.095 | 0.075 |
| Cysteine (C) | 0.032 | 0.022 | 0.054 | 0.054 | 0.053 | 0.033 | 0.032 | 0.032 | 0 | 0 |
| Aspartic Acid (D) | 0 | 0 | 0 | 0 | 0 | 0.03 | 0.027 | 0.027 | 0.071 | 0.075 |
| Glutamic Acid (E) | 0.048 | 0.045 | 0 | 0 | 0 | 0.052 | 0.049 | 0.049 | 0.071 | 0.075 |
| Phenylalanine (F) | 0.032 | 0.034 | 0.054 | 0.054 | 0.053 | 0.061 | 0.065 | 0.065 | 0.071 | 0.075 |
| Glycine (G) | 0.048 | 0.045 | 0.054 | 0.054 | 0.053 | 0.044 | 0.042 | 0.042 | 0.143 | 0.15 |
| Histidine (H) | 0.032 | 0.034 | 0.027 | 0.027 | 0.026 | 0.035 | 0.034 | 0.034 | 0.071 | 0.075 |
| Isoleucine (I) | 0.016 | 0.045 | 0.027 | 0.027 | 0.026 | 0.068 | 0.068 | 0.068 | 0.071 | 0.05 |
| Lysine (K) | 0.065 | 0.067 | 0.027 | 0.027 | 0.026 | 0.04 | 0.042 | 0.042 | 0.048 | 0.05 |
| Leucine (L) | 0.113 | 0.146 | 0.081 | 0.081 | 0.079 | 0.089 | 0.093 | 0.095 | 0.048 | 0.05 |
| Methionine (M) | 0 | 0.011 | 0 | 0 | 0 | 0.035 | 0.034 | 0.034 | 0.024 | 0.025 |
| Asparagine (N) | 0.129 | 0.101 | 0.162 | 0.162 | 0.158 | 0.054 | 0.053 | 0.053 | 0.024 | 0.025 |
| Proline (P) | 0.032 | 0.034 | 0 | 0 | 0 | 0.047 | 0.057 | 0.055 | 0 | 0 |
| Glutamine (Q) | 0.032 | 0.034 | 0.027 | 0.027 | 0.026 | 0.042 | 0.04 | 0.04 | 0.024 | 0.025 |
| Arginine (R) | 0.065 | 0.045 | 0.027 | 0.027 | 0.026 | 0.047 | 0.049 | 0.049 | 0.024 | 0.025 |
| Serine (S) | 0.081 | 0.079 | 0.135 | 0.135 | 0.132 | 0.042 | 0.042 | 0.042 | 0.048 | 0.05 |
| Threonine (T) | 0.081 | 0.067 | 0.135 | 0.135 | 0.132 | 0.052 | 0.055 | 0.055 | 0 | 0 |
| Valine (V) | 0.081 | 0.09 | 0.054 | 0.054 | 0.053 | 0.077 | 0.074 | 0.074 | 0.143 | 0.15 |
| Tryptophan (W) | 0 | 0 | 0 | 0 | 0 | 0.03 | 0.027 | 0.027 | 0 | 0 |
| Tyrosine (Y) | 0.032 | 0.022 | 0.027 | 0 | 0.026 | 0.056 | 0.055 | 0.055 | 0.024 | 0.025 |

Table 3: Hydrophobic and Hydrophilic residues content computed by CLC Workbench

| ID number | AAA35524.1 | NP_000406.1 | 3HGZ E | 2KB8 A | 2L86 A | AAC50301.1 | AAC50300.1 | NP_001733.1 | 2WK3 D | 2LMP K |
|---|------------|-------------|--------|--------|--------|------------|------------|-------------|--------|--------|
| Counts of Hydrophobic residues(A,F,G,I,L,M,P,V,W) | 25 | 43 | 14 | 14 | 14 | 221 | 248 | 248 | 25 | 23 |
| Counts of Hydrophilic residues(C,N,Q,S,T,Y) | 24 | 29 | 20 | 19 | 20 | 119 | 131 | 131 | 5 | 5 |

Table 4: Parameters computed by Expsy ProtParam

| ID no | PI | Mol wt | -R | +R | EC | II | AI | GRAVY |
|-------------|------|---------|----|----|--------|-------|--------|--------|
| AAA35524.1 | 9.78 | 6875.8 | 3 | 8 | 3105 | 23.85 | 81.77 | -0.456 |
| NP_000406.1 | 9.83 | 9806.4 | 4 | 10 | 3105 | 26.96 | 108.43 | 0.035 |
| 3HGZ E | 8.9 | 3906.3 | 0 | 2 | 1615 | 1.94 | 68.65 | -0.097 |
| 2KB8 A | 8.96 | 3854.4 | 0 | 2 | 125 | 1.67 | 68.65 | -0.062 |
| 2L86 A | 8.9 | 4017.6 | 0 | 2 | 1615 | 1.89 | 66.84 | -0.095 |
| AAC50301.1 | 7.88 | 49882 | 35 | 37 | 108135 | 42.81 | 90.16 | 0.041 |
| AAC50300.1 | 8.18 | 47171.9 | 32 | 35 | 103540 | 43.84 | 93.84 | 0.105 |
| NP_001733.1 | 8.6 | 55344.5 | 36 | 43 | 111115 | 44.29 | 91.1 | 0.061 |
| 2WK3 D | 5.31 | 4514.1 | 6 | 3 | 1490 | 18.17 | 97.38 | 0.205 |
| 2LMP K | 5.31 | 4329.8 | 6 | 3 | 1490 | 18.58 | 90 | 0.057 |

Table 5: Representation of helix, sheet, turn, coils by through online tool SOPMA

| | ID no | AAA35524.1 | NP_000406.1 | 3HGZ E | 2KB8 A | 2L86 A | AAC50301.1 | AAC50300.1 | NP_001733.1 | 2WK3 D | 2LMP K |
|----------|----------------|------------|-------------|--------|--------|--------|------------|------------|-------------|--------|--------|
| Helix(H) | Residue totals | 22 | 41 | 10 | 10 | 10 | 156 | 141 | 171 | 2 | 2 |
| | Percentage% | 35.48 | 46.07 | 27.03 | 26.32 | 26.32 | 36.53 | 34.9 | 36.08 | 4.76 | 5 |
| Sheet(E) | Residue totals | 12 | 15 | 8 | 9 | 9 | 114 | 114 | 125 | 22 | 19 |
| | Percentage% | 19.35 | 16.85 | 21.62 | 23.68 | 23.68 | 26.7 | 28.22 | 26.37 | 52.38 | 47.5 |
| Turn(T) | Residue totals | 2 | 3 | 4 | 4 | 4 | 44 | 41 | 42 | 8 | 7 |
| | Percentage% | 3.23 | 3.37 | 10.81 | 10.53 | 10.53 | 10.3 | 10.15 | 8.86 | 19.05 | 17.5 |
| Coils(C) | Residue totals | 26 | 30 | 15 | 15 | 15 | 113 | 108 | 136 | 10 | 12 |
| | Percentage% | 41.94 | 33.71 | 40.54 | 39.47 | 39.47 | 26.46 | 26.73 | 28.69 | 23.81 | 30 |

Table 6: SMART analysis of Amylin protein sequences

| ID number | Start | End | E -value |
|-------------|-------|-----|-----------|
| AAA35524.1 | 5 | 47 | 5.47e-17 |
| NP_000406.1 | 32 | 74 | 5.47e-17 |
| 3HGZ E | 1 | 37 | 8.77e-7 |
| 2KB8 A | 1 | 36 | 0.0000198 |
| 2L86 A | 1 | 37 | 8.77e-7 |
| AAC50301.1 | 21 | 96 | 7.49e-27 |
| AAC50300.1 | 1 | 73 | 4.72e-24 |
| NP_001733.1 | 68 | 143 | 7.49e-27 |
| 2WK3 D | | | No domain |
| 2LMP K | | | No domain |

Table 7: SVMprot analysis of Amylin protein sequences

| Accession number | Protein family name | | | | | | | | | | | | | | | | | | | |
|--|---------------------|---------|-------------|---------|---------|---------|---------|---------|---------|---------|------------|---------|------------|---------|-------------|---------|---------|---------|---------|---------|
| | AAA35524.1 | | NP_000406.1 | | 3HGZ E | | 2KB8 A | | 2L86 A | | AAC50301.1 | | AAC50300.1 | | NP_001733.1 | | 2WK3 D | | 2LMP K | |
| | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value | R Value | P Value |
| Iron binding | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| All DNA binding | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Photoreceptor | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Magnesium binding | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Hormone | NA | NA | 6 | 99 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Transmembrane | NA | NA | 1.2 | 65 | NA | NA | NA | NA | NA | NA | 6.8 | 99 | 10 | 99 | 10 | 99 | NA | NA | NA | NA |
| All lipid binding proteins | NA | NA | 1.1 | 62 | NA | NA | NA | NA | NA | NA | 2.1 | 85 | 2.8 | 93 | 2.8 | 93 | NA | NA | NA | NA |
| Outer membrane | NA | NA | 1 | 59 | NA | NA | NA | NA | NA | NA | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA |
| Metal binding | NA | NA | 1 | 59 | NA | NA | NA | NA | NA | NA | 1 | 59 | 1.1 | 62 | 1.1 | 62 | NA | NA | NA | NA |
| Calcium binding | NA | NA | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | 1 | 59 | 1 | 59 | NA | NA | NA | NA |
| TC 1 C channels/pores | NA | NA | 1 | 59 | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| G-Protein Coupled Receptors | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | 4.4 | 98 | 8.5 | 99 | 8.5 | 99 | NA | NA | NA | NA |
| 7 Transmembrane receptor | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | 1.2 | 65 | 1 | 59 | 1 | 59 | NA | NA | NA | NA |
| TC 1 A channels/pores | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | 1.1 | 62 | 1.2 | 65 | 1.2 | 65 | NA | NA | NA | NA |
| 7 Transmembrane receptor (secretin family) | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | 1 | 59 | 1 | 59 | 1 | 59 | NA | NA | NA | NA |
| Chlorophyll biosynthesis | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | NA | 1 | 59 | 1 | 59 | NA | NA | NA | NA |

(R value and P values are in percentage)

Table 8: RNA structure stems with free energy (ref/NP_000406.1)

| Stem no | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 |
|-----------------------|-------|-----|-----|------|------|------|------|------|------|------|------|
| Free energy(Kkal/mol) | -15.8 | -13 | -10 | -9.4 | -8.7 | -8.4 | -8.2 | -7.7 | -6.9 | -6.2 | -6.2 |

Domain architecture analysis: Domain organization and domain composition was analyzed using Simple Modular Architecture Research Tool (SMART) (Table 6).

SVMprot analysis: SVMprot tool was used to predict protein function, and to classify the distantly related proteins (Table 7).

Trans-membrane region prediction: TMpred was used to predict transmembrane helices. The TMpred software is available through internet access (http://www.ch.embnet.org/software/TMPRED_form.html) (Fig 1).

Sequence Homology Analysis: The sequence homology was analyzed by MUSCLE (<http://www.ebi.ac.uk/Tools/msa/muscle/>) (Fig 2).

RNA structure prediction: The protein sequences were reverse transcribed to DNA using Sequence Manipulation Suite (SMS) (http://www.bioinformatics.org/sms2/rev_trans.html). The reverse transcribed DNA was converted to RNA using transcriptional and translational tool (<http://www.attotron.com/cybertory/analysis/trans.html>). RNA structure was predicted using (http://www.genebee.msu.su/services/rna2_reduced.html) (Fig 3).

Swiss model: Homology-modeling was performed using SWISS-MODEL accessible via the ExPasy web server. (<http://swissmodel.expasy.org/>) (Fig 4)

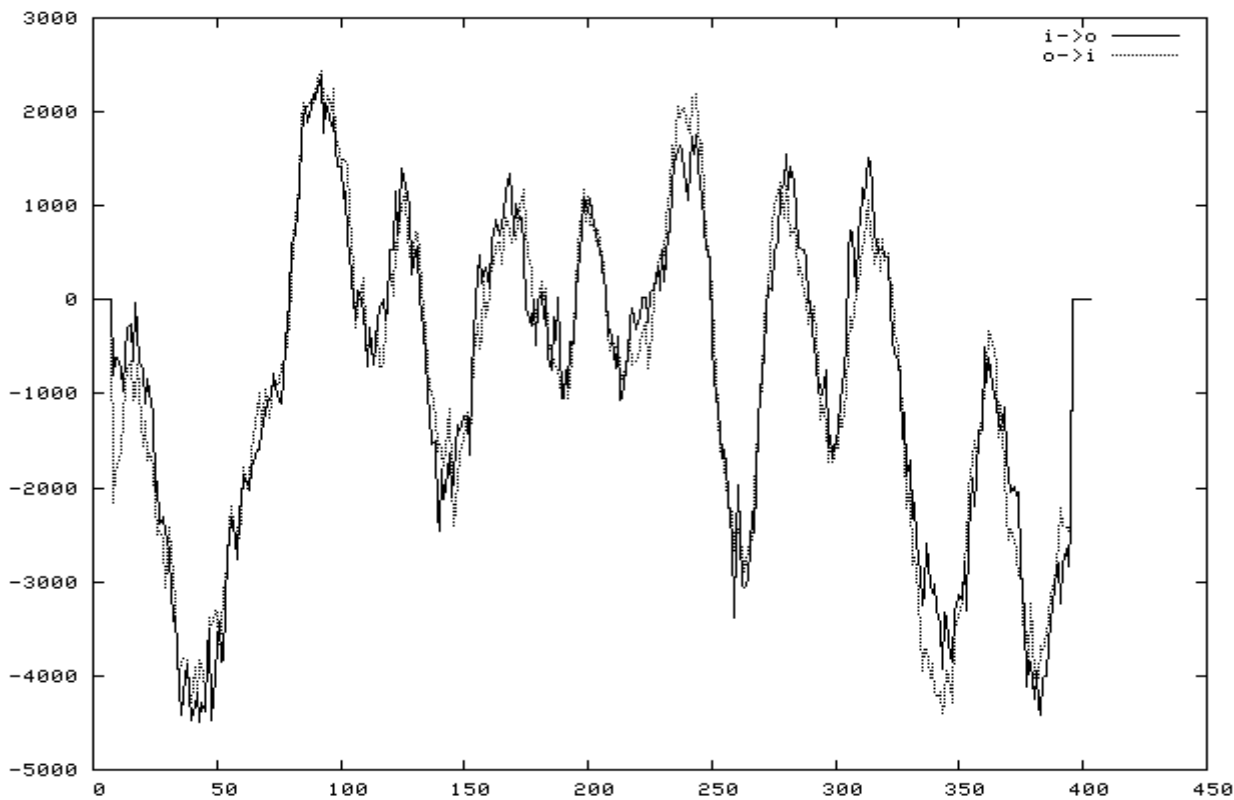
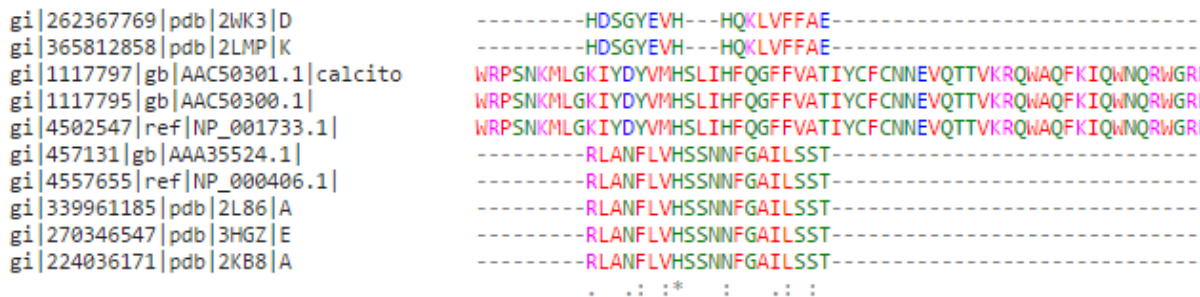


Fig.1 TMpred graph: (ref AAC50300.1)



Identity (*): Strongly similar (:): Weakly similar (.)

Fig.2 Multiple sequence alignment of Amylin protein sequence by MUSCLE software

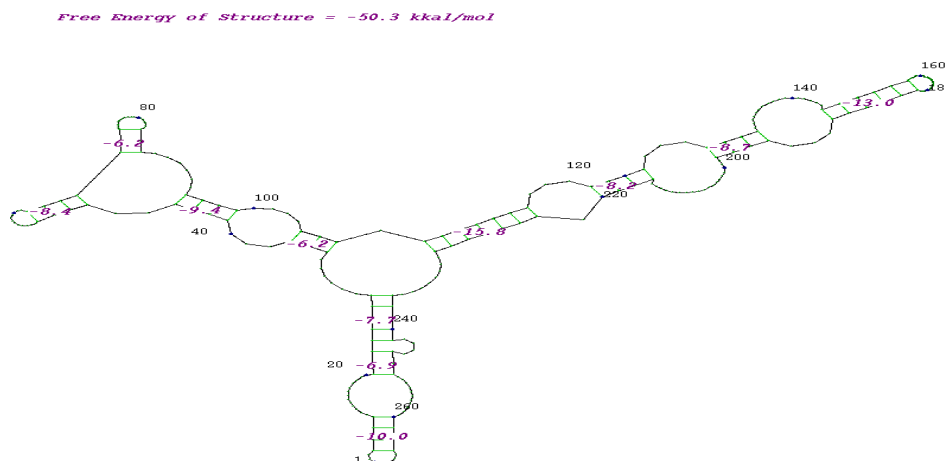


Fig.3 RNA structure prediction: (AAC50300.1)

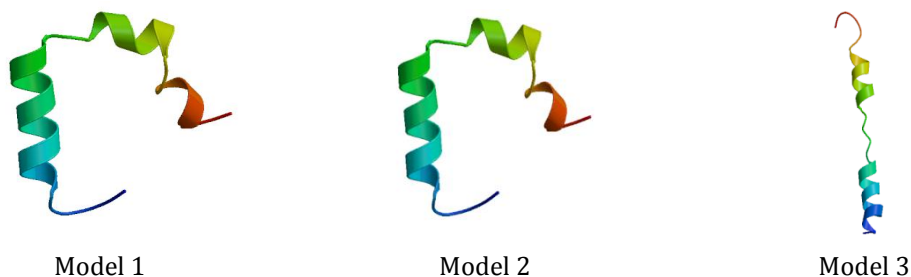
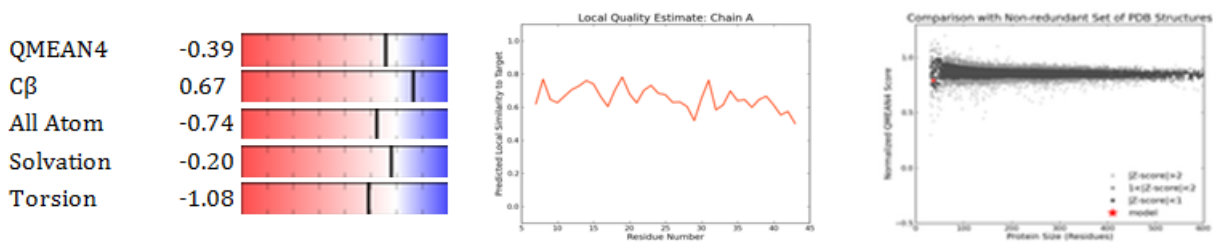


Fig.4 SWISS MODEL: (AAA35524.1)

| Model #01 | File | Built with | Oligo-State | Ligands | GMQE | QMEAN4 |
|-----------|---------------------|----------------------|-------------|---------|------|--------|
| | PDB | ProMod Version 3.70. | MONOMER | None | 0.68 | -0.39 |



| Template | Seq Identity | Oligo-state | Found by | Method | Resolution | Seq Similarity | Range | Coverage | Description |
|--------------------------|--------------|-------------|----------|--------|------------|----------------|--------|----------|---------------------------|
| 2l86.1.A | 100.00 | monomer | BLAST | NMR | NA | 0.61 | 7 - 43 | 0.60 | Islet amyloid polypeptide |

The template contained no ligands.

Target HQVEKRRKNCNTATCATQRLANFLVHSSNFFGAILSSTNVGSNTYGKRNAVEVLKREPLNYLPL
 2l86.1.A -----KCNTATCATQRLANFLVHSSNFFGAILSSTNVGSNTY-----

Fig.5 Model building using SWISS MODEL (AAA35524.1)



Fig.6 3D view of the structure with ligand binding: (AAA35524.1)

Conclusion

Amylin, a naturally occurring hormone, is a normal product of beta cells and is co-released with insulin. In the present work, we have addressed the amino acid composition, physiochemical properties, structure and function of amyloid fibrils by using a domain of the diabetes-associated IAPP peptides as model.

Amyloid fibrils formation varies greatly with environmental conditions like temperature, pH, oxidative stress, ionic strength, peptide concentration etc. These highly ordered protein fibrils form aggregates and deposition of which leads to several degenerative diseases such as Alzheimer’s disease, Parkinson’s disease, Type II diabetes and so on. The proteomic analysis and characterization of amylin is

done so that a complete knowledge of amylin protein is determined and a site of interest which is to be treated can be identified easily and novel drug can be induced. Usage of bio informatics tool gives a complete knowledge to treat type 2 diabetes which is associated with formation of amyloids from amylin protein.

The understanding of their physiochemical characteristics, function, structure and their mechanism of action can serve as a key therapeutic approach towards treating amyloid diseases.

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